

Organic Syntheses

Instructions for Authors

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Editor in Chief

All organic chemists have experienced frustration at one time or another when attempting to repeat reactions based on experimental procedures found in journal articles. To ensure reproducibility, *Organic Syntheses* requires experimental procedures written with considerably more detail as compared to the typical procedures found in other journals and in the "Supporting Information" sections of papers. In addition, each *Organic Syntheses* procedure is carefully "checked" for reproducibility in the laboratory of a member of the Board of Editors.

Even with these more detailed procedures, the experience of *Organic Syntheses* editors is that difficulties often arise in obtaining the results and yields reported by the submitters of procedures. To expedite the checking process and ensure success, we have prepared the following "Instructions for Authors" as well as a [Checklist for Authors](#) and [Characterization Checklist](#) to assist you in confirming that your procedure conforms to these requirements. Please include a completed Checklist together with your procedure at the time of submission. Procedures submitted to *Organic Syntheses* will be carefully reviewed upon receipt and procedures lacking any of the required information will be returned to the submitters for revision.

Scale and Optimization

The appropriate scale for procedures will vary widely depending on the nature of the chemistry and the compounds synthesized in the procedure. However, some general guidelines are possible. For procedures in which the principal goal is to illustrate a synthetic method or strategy, it is expected, in general, that the procedure should result in **at least 2 g** and no more than 50 g of the final product. In cases where the point of the procedure is to provide an efficient method for the preparation of a useful reagent or synthetic building block, the appropriate scale also should be between 2 and 50 g of final product. Exceptions to these guidelines may be granted in special circumstances. For example, procedures describing the preparation of reagents employed as catalysts will often be acceptable on a scale of less than 2 g.

In considering the scale for an *Organic Syntheses* procedure, authors should also take into account the cost of reagents and starting materials. In general, the Editors will not accept procedures for checking in which the cost of any one of the reactants exceeds **\$500** for a single full-scale run. Authors are required to identify the most expensive reagent or starting material on the procedure submission checklist and to estimate its cost per run of the procedure. Authors should purchase chemicals from common suppliers that are well known to the chemistry community.

It is expected that all aspects of the procedure will have been optimized by the authors prior to submission, and it is required that each reaction will have been carried out at least twice on exactly the scale described in the procedure, and with the results reported in the manuscript.

It is appropriate to report the weight, yield, and purity of the product of each step in the procedure as a range. In any case where a reagent is employed in significant excess, a Note should be included explaining why an excess of that reagent is necessary. If possible, the Note should indicate the effect of using amounts of reagent less than that specified in the procedure.

The Checking Process

A unique feature of papers published in *Organic Syntheses* is that each procedure and all characterization data is carefully checked for reproducibility in the laboratory of a member of the Board of Editors. In the event that an editor finds it necessary to make any modifications in an experimental procedure, then the published article incorporates the modified procedure, with an explanation and mention of the original protocol often included in a Note. The yields reported in the published article are always those obtained by the checkers. In general, the characterization data in the published article also is that of the checkers, unless there are significant differences with the data obtained by the authors, in which case the author's data will also be reported in a Note.

Reaction Apparatus

Describe the size and type of flask (number of necks) and indicate how every neck is equipped.

"A 500-mL, three-necked, round-bottomed flask equipped with an 3-cm Teflon-coated magnetic stirbar, a 250-mL pressure-equalizing addition funnel fitted with an argon inlet, and a rubber septum is charged with"

Indicate how the reaction apparatus is dried and whether the reaction is conducted under an inert atmosphere. Note that in general balloons are not acceptable as a means of maintaining an inert atmosphere unless warranted by special circumstances. For reactions

conducted below -20 °C it is advisable to include a thermocouple or thermometer to monitor the internal temperature of the reaction mixture.

Further details concerning the reaction apparatus can be incorporated in the text of the procedure or included in a Note.

“The apparatus is flame-dried and maintained under an atmosphere of argon during the course of the reaction.”

Photographs are required depicting the reaction apparatus and in the case of procedures involving unusual glassware or especially complicated reaction setups, authors may wish to include a drawing of the apparatus in the text or in a Note (for example, see *Org. Synth.* **2016**, 93, 127-145).

Use of Gloveboxes

When a glovebox is employed in a procedure, justification must be provided in a Note and the consequences of carrying out the operation without using a glovebox must be discussed.

Reagents and Starting Materials

All chemicals employed in the procedure must be commercially available or described in an earlier *Organic Syntheses* or *Inorganic Syntheses* procedure. For other compounds, a procedure should be included either as one or more steps in the text or, in the case of relatively straightforward preparations of reagents, as a Note. In the latter case, all requirements with regard to characterization, style, and detail also apply. Authors are encouraged to consult with the Associate Editor if they have any question as to whether to include such steps as part of the text or as a Note.

Authors are encouraged to consider the use of “substitute solvents” in place of more hazardous alternatives. For example, the use of *t*-butyl methyl ether (MTBE) should be considered as a substitute for diethyl ether, particularly in large scale work. Authors are referred to the articles “Sanofi’s Solvent Selection Guide: A Step Toward More Sustainable Processes” (Prat, D.; Pardigon, O.; Flemming, H.-W.; Letestu, S.; Ducandas, V.; Isnard, P.; Guntrum, E.; Senac, T.; Ruisseau, S.; Cruciani, P. Hosek, P. *Org. Process Res. Dev.* **2013**, 17, 1517-1525) and “Solvent Replacement for Green Processing” (Sherman, J.; Chin, B.; Huibers, P. D. T.; Garcia-Valis, R.; Hatton, T. A. *Environ. Health Perspect.* **1998**, 106 (Supplement I, 253-271) as well as the references cited therein for discussions of this subject. In addition, a link to a “solvent selection guide” can be accessed via the American Chemical Society Green Chemistry website at <http://www.acs.org/content/acs/en/greenchemistry/research-innovation/tools-for-green-chemistry.html>.

In one or more Notes, indicate the purity or grade of each reagent, solvent, etc. It is also expected that authors will indicate the source (company the chemical was purchased from),

particularly in the case of chemicals where it is suspected that the composition (trace impurities, etc.) may vary from one supplier to another. In cases where reagents are purified, dried, "activated" (e.g., Zn dust), etc., a detailed description of the procedure used should be included in a Note. In other cases, indicate that the chemical was "used as received".

"Diisopropylamine (99.5%) was obtained from Aldrich Chemical Co., Inc. and distilled under argon from calcium hydride before use. THF (99+%) was obtained from Mallinckrodt, Inc. and distilled from sodium benzophenone ketyl. Diethyl ether (99.9%) was purchased from Aldrich Chemical Co., Inc. and purified by pressure filtration under argon through activated alumina. Methyl iodide (99%) was obtained from Aldrich Chemical Co., Inc. and used as received."

The amount of each reactant must be provided in parentheses in the order mL, g, mmol, and equivalents with careful consideration to the correct number of **significant figures**. Avoid indicating amounts of reactants with more significant figures than makes sense. For example, "437 mL of THF" implies that the amount of solvent must be measured with a level of precision that is unlikely to affect the outcome of the reaction. Likewise, "5.00 equiv" implies that an amount of excess reagent must be controlled to a precision of 0.01 equiv.

The concentration of solutions should be expressed in terms of molarity or normality, and not percent (e.g., 1 N HCl, 6 M NaOH, not "10% HCl").

Reaction Procedure

Describe every aspect of the procedure clearly and explicitly. Indicate the order of addition and time for addition of all reagents and how each is added (via syringe, addition funnel, etc.).

Indicate the temperature of the reaction mixture (preferably internal temperature). Describe the type of cooling (e.g., "dry ice-acetone bath") and heating (e.g., oil bath, heating mantle) methods employed. Be careful to describe clearly all cooling and warming cycles, including initial and final temperatures and the time interval involved.

Describe the appearance of the reaction mixture (color, homogeneous or not, etc.) and describe all significant changes in appearance during the course of the reaction (color changes, gas evolution, appearance of solids, exotherms, etc.).

Indicate how the reaction can be monitored to determine the extent of conversion of reactants to products. In the case of reactions monitored by TLC, provide details in a Note, including eluent, R_f values, and method of visualization. For reactions followed by GC, HPLC, or NMR analysis, provide details on analysis conditions and relevant diagnostic peaks.

"The progress of the reaction was followed by TLC analysis on silica gel with 1:4 EtOAc-hexane as eluent and visualization with *p*-anisaldehyde. The ketone starting material has $R_f = 0.40$ (green) and the alcohol product has $R_f = 0.25$ (blue)."

Reaction Workup

Details should be provided for reactions in which a “quenching” process is involved. Describe the composition and volume of quenching agent, and time and temperature for addition. In cases where reaction mixtures are added to a quenching solution, be sure to also describe the setup employed.

“The resulting mixture was stirred at room temperature for 15 h, and then carefully poured over 10 min into a rapidly stirred, ice-cold aqueous solution of 1 N HCl in a 500-mL Erlenmeyer flask equipped with a magnetic stirbar.”

For extractions, the number of washes and the volume of each should be indicated as well as the size of the separatory funnel.

For concentration of solutions after workup, indicate the method and pressure and temperature used.

“The reaction mixture is diluted with 200 mL of water and transferred to a 500-mL separatory funnel, and the aqueous phase is separated and extracted with three 100-mL portions of ether. The combined organic layers are washed with 75 mL of water and 75 mL of saturated NaCl solution, dried over 25 g of MgSO₄, filtered through a 250-mL medium porosity sintered glass funnel, and concentrated by rotary evaporation (25 °C, 20 mmHg) to afford 3.25 g of a yellow oil.”

“The solution is transferred to a 250-mL, round-bottomed flask equipped with a magnetic stirbar and a 15-cm Vigreux column fitted with a short path distillation head, and then concentrated by careful distillation at 50 mmHg (bath temperature gradually increased from 25 to 75 °C).”

In cases where solid products are filtered, describe the type of filter funnel used and the amount and composition of solvents used for washes.

“ . . . and the resulting pale yellow solid is collected by filtration on a Büchner funnel and washed with 100 mL of cold (0 °C) hexane.”

When solid or liquid compounds are dried under vacuum, indicate the pressure employed (rather than stating “reduced pressure” or “dried *in vacuo*”).

“ and concentrated at room temperature by rotary evaporation (20 mmHg) and then at 0.01 mmHg to provide ”

“The resulting colorless crystals are transferred to a 50-mL, round-bottomed flask and dried overnight in a 100 °C oil bath at 0.01 mmHg.”

Purification: Distillation

Describe distillation apparatus including the size and type of distillation column. Indicate temperature (and pressure) at which all significant fractions are collected.

“ . . . and transferred to a 100-mL, round-bottomed flask equipped with a magnetic stirbar. The product is distilled under vacuum through a 12-cm, vacuum-jacketed column of glass helices (Note 16) topped with a Perkin triangle. A forerun (ca. 2 mL) is collected and discarded, and the desired product is then obtained, distilling at 50-55 °C (0.04-0.07 mmHg) . . . ”

Purification: Column Chromatography

Provide information on TLC analysis in a Note, including eluent, R_f values, and method of visualization.

Provide dimensions of column and amount of silica gel used; in a Note indicate source and type of silica gel.

Provide details on eluents used, and number and size of fractions.

“The product is charged on a column (5 x 10 cm) of 200 g of silica gel (Note 15) and eluted with 250 mL of hexane. At that point, fraction collection (25-mL fractions) is begun, and elution is continued with 300 mL of 2% EtOAc-hexane (49:1 hexanes:EtOAc) and then 500 mL of 5% EtOAc-hexane (19:1 hexanes:EtOAc). The desired product is obtained in fractions 24-30, which are concentrated by rotary evaporation (25 °C, 15 mmHg)”

Use of Automated Column Chromatography

Automated column chromatography should not be used for purification of products unless the use of such systems is essential to the success of the procedure. When automated column chromatography equipment is employed in a procedure, justification must be provided in a Note and the consequences of carrying out the purification using conventional column chromatography must be discussed.

Purification: Recrystallization

Describe procedure in detail. Indicate solvents used (and ratio of mixed solvent systems), amount of recrystallization solvents, and temperature protocol. Describe how crystals are isolated and what they are washed with. A photograph of the crystalline product is often valuable to indicate the form and color of the crystals.

“The solid is dissolved in 100 mL of hot diethyl ether (30 °C) and filtered through a Buchner funnel. The filtrate is allowed to cool to room temperature, and 20 mL of hexane is added. The solution is cooled at -20 °C overnight and the resulting crystals are collected by suction filtration on a Buchner funnel, washed with 50 mL of ice-cold hexane, and then transferred to a 50-mL, round-bottomed flask and dried overnight at 0.01 mmHg to provide”

Characterization and Proof of Purity

Physical properties of the product such as color, appearance, crystal forms, melting point, etc. should be included in the text of the procedure. Comments on the stability of the product to storage, etc. should be provided in a Note.

In a Note, provide data establishing the identity of the product. This will generally include IR, MS, ¹H-NMR, and ¹³C-NMR data, and in some cases UV data. Copies of the proton and carbon NMR spectra for the products of each step in the procedure should be submitted showing integration for all resonances. Submission of copies of the NMR spectra for other nuclei are encouraged as appropriate.

In the same Note, provide analytical data establishing that the purity of the **isolated** product is at least 97%. **Note that this data should be obtained for the material on which the yield of the reaction is based**, not for a sample that has been subjected to additional purification by chromatography, distillation, or crystallization. Elemental analysis for carbon and hydrogen (and nitrogen if present) agreeing with calculated values within 0.4% is preferred. However, **quantitative** NMR, GC, or HPLC analyses involving measurements versus an internal standard will also be accepted. See Appendix I of these instructions for procedures for quantitative analysis of purity by NMR and chromatographic methods. Authors must provide details on equipment and conditions for GC and HPLC analyses, and copies of NMR spectra and chromatograms used in the quantitative analyses must be submitted with articles for publication.

In procedures involving non-racemic, enantiomerically enriched products, optical rotations should generally be provided, but **enantiomeric purity must be determined by another method** such as chiral HPLC or GC analysis.

In cases where the product of one step is used without purification in the next step, a Note should be included describing how a sample of the product can be purified and providing characterization data for the pure material. Copies of the proton NMR spectra of both the product both *before* and *after* purification should be submitted.

Safety Note and Hazard Warnings

Effective in August 2017, the first Note in every article is devoted to addressing the safety aspects of the procedures described in the article. The Article Template provides the

required wording and format for Note 1, which reminds readers of the importance of carrying out risk assessments and hazard analyses prior to performing all experiments:

Prior to performing each reaction, a thorough hazard analysis and risk assessment should be carried out with regard to each chemical substance and experimental operation on the scale planned and in the context of the laboratory where the procedures will be carried out. Guidelines for carrying out risk assessments and for analyzing the hazards associated with chemicals can be found in references such as Chapter 4 of "Prudent Practices in the Laboratory" (The National Academies Press, Washington, D.C., 2011; the full text can be accessed free of charge at http://www.nap.edu/catalog.php?record_id=12654). See also "Identifying and Evaluating Hazards in Research Laboratories" (American Chemical Society, 2015) which is available via the associated website "Hazard Assessment in Research Laboratories" at <https://www.acs.org/content/acs/en/about/governance/committees/chemicalsafety/hazard-assessment.html>. In the case of this procedure, the risk assessment should include (but not necessarily be limited to) an evaluation of the potential hazards associated with (*enter list of chemicals here*), as well as the proper procedures for (*list any unusual experimental operations here*). (*Provide additional cautions with regard to exceptional hazards here*).

For the required list of chemicals, authors should include all reactants, solvents, and other chemicals involved in the reactions described in the article.

With regard to the list of experimental operations, this list should be limited to those operations that potentially pose significant hazards. Examples may include

- Vacuum distillations
- Reactions run at elevated pressure or in sealed reaction vessels
- Photochemical reactions

In the case of experiments that involve exceptional hazards such as the use of pyrophoric or explosive substances, and substances with a high degree of acute or chronic toxicity, authors should provide additional guidelines for how to carry out the experiment so as to minimize risk. These instructions formerly would have appeared as red "Caution Notes" in *Organic Syntheses* articles. Note that it is not essential to describe general safety procedures such as working in a hood, avoiding skin contact, using eye protection, etc., since these are discussed in the Prudent Practices reference mentioned in the "Working with Hazardous Chemicals" statement within each article.

Efforts should be made to avoid the use of toxic and hazardous solvents and reagents when less hazardous alternatives are available.

Discussion Section

The style and content of the discussion section will depend on the nature of the procedure.

For procedures that provide an improved method for the preparation of an important reagent or synthetic building block, the discussion should focus on the advantages of the new

approach and should describe and reference all of the earlier methods used to prepare the title compound.

In the case of procedures that illustrate an important synthetic method or strategy, the discussion section should provide a mini-review on the new methodology. The scope and limitations of the method should be discussed, and it is generally desirable to include a table of examples. Please be sure each table is numbered and has a title. Competing methods for accomplishing the same overall transformation should be described and referenced. A brief discussion of mechanism may be included if this is useful for understanding the scope and limitations of the method.

Titles of Articles

In cases where the main thrust of the article is the illustration of a synthetic method of general utility, the title of the article should incorporate reference to that method. Inclusion of the name of the final product is acceptable but not required. In the case of articles where the objective is the preparation of a specific compound of importance (such as a chiral ligand), then the name of that compound should be part of the title.

Examples

Title without name of product:

“Stereoselective Synthesis of 3-Arylacrylates by Copper-Catalyzed Syn Hydroarylation” (*Org. Synth.* **2010**, 87, 53).

Title including name of final product (note name of product is not required):

“Catalytic Enantioselective Borane Reduction of Benzyl Oximes: Preparation of (S)-1-Pyridin-3-yl-ethylamine Bis Hydrochloride” (*Org. Synth.* **2010**, 87, 36).

Title where preparation of specific compound is the subject:

“Preparation of (S)-3,3'-Bis-Morpholinomethyl-5,5',6,6',7,7',8,8'-octahydro-1,1'-bi-2-naphthol” (*Org. Synth.* **2010**, 87, 59).

Heading Scheme

The title of the article should be followed by a “Heading Scheme” comprising separate equations for each step in the article. Authors should consult the article template for instructions concerning ChemDraw settings and format. In general, reaction equations should not include details such as reaction time and the number of equivalents of reagents, with the exception of reactants employed in catalytic amounts which can be labeled as “cat.” or by specifying mol%.

Style and Format for Text

The text of articles should follow the style guidelines used for organic chemistry articles published in the ACS journals such as *J. Am. Chem. Soc.*, *J. Org. Chem.*, *Org. Lett.*, etc. as described in the ACS Style Guide (3rd Ed.). Past tense should be used in the description of procedures. The text of the procedure should be created using the Word template available on the *Organic Syntheses* website. Specific instructions with regard to the manuscript format (font, spacing, margins) is available on the website in the "Instructions for Article Template" and embedded within the Article Template itself.

Style and Format for Tables and Schemes

Chemical structures and schemes should be drawn using the standard ACS drawing parameters (in ChemDraw, the parameters are found in the "ACS Document 1996" option) with a maximum full size width of 15 cm (5.9 inches). The graphics files should then be pasted into the Word document at the correct location and the size reduced to 75% using "Format Picture" (Mac) or "Size and Position" (Windows). Graphics files must also be submitted separately. All Tables that include structures should be entirely prepared in the graphics (ChemDraw) program and inserted into the word processing file at the appropriate location. Tables that include multiple, separate graphics files prepared in the word processing program will require modification.

Tables and schemes should be numbered and should have titles. The title for a Table should be included immediately above the table. The title for a scheme should be placed immediately below the scheme. Use 10 point Palatino Bold font in the ChemDraw file for all titles. For footnotes in Tables use Helvetica (or Arial) 9 point font and place these immediately below the Table.

Photographs

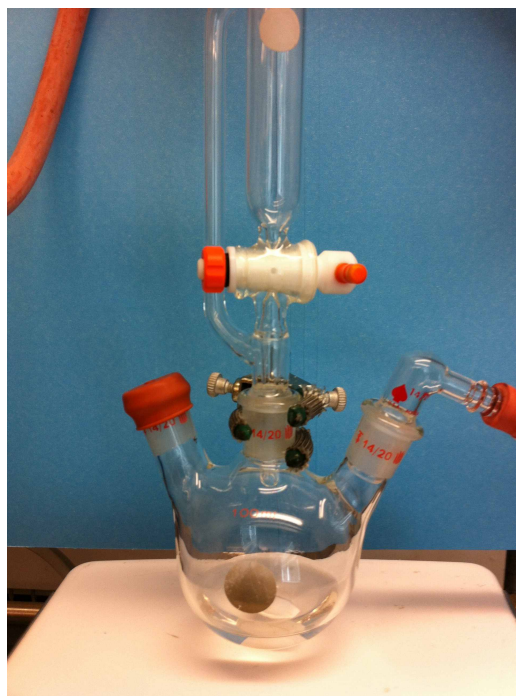
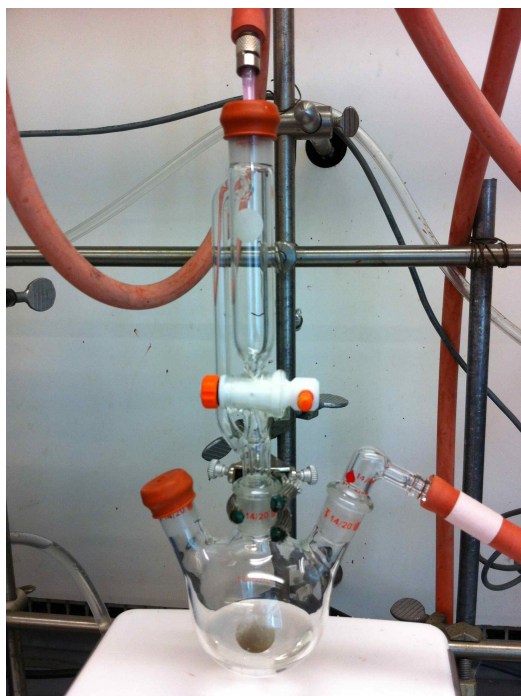
Photographs illustrating key elements of procedures are required in every article published in *Organic Syntheses*. Authors are expected to furnish photos with their original submissions and photos may also be provided by the Checkers of procedures. Photographs should be inserted into articles at the place in the text and Notes where they are first referred to and should be numbered and labeled as Figures with descriptive titles. Particularly useful subjects for photographs include:

- Photos of reaction flasks depicting how each neck is equipped
- Photos of reaction mixtures illustrating color changes, heterogeneity, etc.
- Photos of TLC plates showing degree of resolution and the color of spots
- Photos of crystalline reaction products illustrating color and crystal type

Photographs with satisfactory resolution can be obtained using phone cameras as well as regular digital cameras. Acceptable formats for submitted photos are jpg and png. The following considerations should be kept in mind in order to obtain the best photographs.

- Use cropped images and closeups to focus attention on the subject of interest.
- Take care with lighting to avoid reflections on glassware that obscure the contents.
- Avoid complicated and distracting backgrounds by placing blank paper or cardboard behind the subject.

For example, in the photographs below, the photo on the right provides a superior view of the reaction setup as compared to the photo on the left.



Acknowledgments and Author's Contact Information

Contact information (institution where the work was carried out and mailing address for the principal author) should be included as footnote 1. This footnote should also include the email address for the principal author. Acknowledgment of financial support should be included in footnote 1.

Biographies and Photographs of Authors

Photographs and 100-word biographies of all authors should be submitted as separate files at the time of the submission of the procedure. The format of the biographies should be similar

to those in recent articles published on the orgsyn.org website. Photographs can be accepted in a number of electronic formats, including tiff and jpeg formats.

Appendix I

Determination of Purity by Quantitative Analysis

Prepared with the assistance of Dr. Margaret Faul and Dr. Christopher Borths, Process Development, Amgen Inc. and Dr. Chris Senanayake, Chemical Development US, Boehringer Ingelheim Pharmaceuticals, Inc.

Introduction

Common techniques for determining the purity of reaction products by quantitative NMR (QNMR) and quantitative chromatographic analysis (HPLC and GC) are presented in this appendix. Authors are reminded that they must provide details on equipment and conditions for GC and HPLC analyses in a Note in their article, and copies of NMR spectra and chromatograms that are used in quantitative analyses must be submitted with articles for publication.

Quantitative NMR (QNMR)¹

A quick and relatively accurate method for the determination of the purity (weight percent) of a compound can be made using proton NMR spectroscopy. A standard compound is used for this determination. Appropriate standards are commercial compounds that can be obtained in high (preferably >99%) purity and that produce a proton NMR signal that is separate and distinct from the signals in the compound being analyzed. Examples of compounds that are available in high purity and are often found useful as internal standards include dimethyl fumarate, ethylene carbonate, and alanine.

To determine the weight percent of the target compound, 10-20 mg of the compound and an approximately equimolar amount of the standard compound (minimum of 10 mg) are accurately weighed into a clean glass container.² The mixture of compounds is completely dissolved in approximately 1 mL of an appropriate NMR solvent such as *d*₆-DMSO, CDCl₃, etc. A proton NMR spectrum is obtained for the mixture with a minimum relaxation delay of 30 s. The weight percent purity of the analyte is calculated by the following equations:

$$\text{Molar ratio} = \frac{\left[\frac{I_{cpd}}{nH_{cpd}} \right]}{\left[\frac{I_{std}}{nH_{std}} \right]}$$
$$\text{wt}\% = \frac{mg_{std} \times MW_{cpd} \times \text{molar ratio} \times P_{std}}{mg_{cpd} \times MW_{std}} \times 100$$

Where,

- Wt% = purity of the sample
- I_{cpd} = proton integral area of a known peak on the compound being analyzed
- nH_{cpd} = number of hydrogens associated with the compound NMR peak
- I_{std} = proton integral area of a known peak on the standard
- nH_{std} = number of hydrogens associated with the standard NMR peak
- mg_{std} = amount in milligrams of the standard compound weighed for analysis
- mg_{cpd} = amount in milligrams of the compound being analyzed
- MW_{std} = molecular weight of the standard
- MW_{cpd} = molecular weight of the compound being analyzed
- P_{std} = wt% purity of the standard expressed as a decimal value. For the purposes of most calculations, a value of 1.00 may be assumed.

Quantitative Chromatographic Analysis

Quantitative analysis by chromatography is typically performed using either an HPLC or GC instrument with an appropriate detector (UV-vis, FID, etc.) that has a linear response range. Once a method is developed that cleanly resolves the peak/compound of interest, it can easily be adapted for quantitative analysis.³ To determine purity, a response factor for the compound of interest must be known, and a standard with known purity (e.g., determined by QNMR) is used to determine this.

The response factor can be calculated from a single-point standard. A single standard sample is analyzed at a concentration within the linear dynamic range of the analytical system and reasonably close to the expected analysis concentration. The response factor is calculated using the following equation:

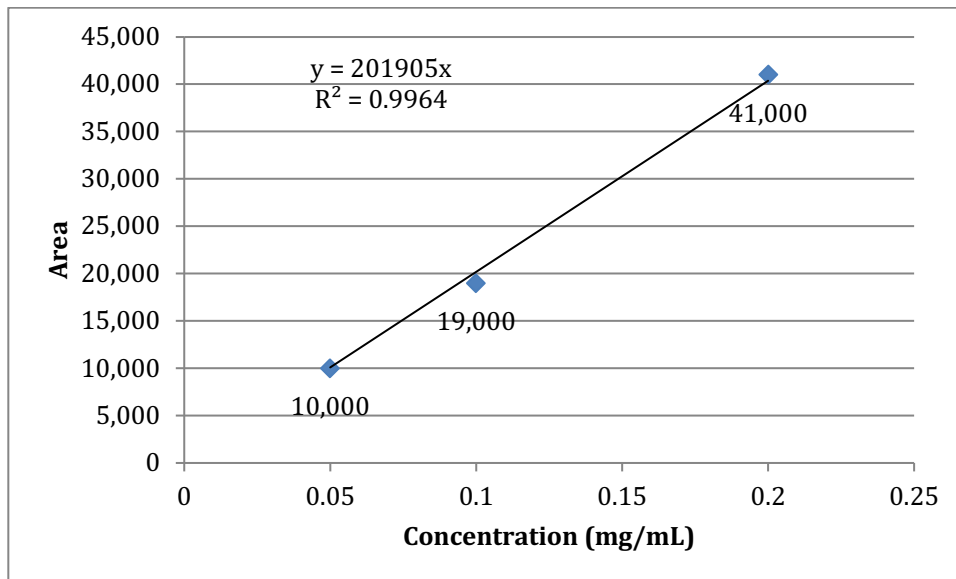
$$RF = \frac{A}{C_{std}}$$

Where,

- RF = response factor
- A = area of the peak of interest
- C_{std} = concentration of the standard sample⁴

Alternately, the response factor can be determined by generating a calibration curve by analyzing standards at multiple concentrations. Using a multi-point calibration curve increases the accuracy and reliability of the concentration measurement by reducing the impact of individual weighing and measurement errors. Standard samples are prepared so that the concentrations are within the linear dynamic range of the detector. These are then analyzed, and the response is plotted against the concentration of each sample. The resulting calibration curve should produce a line. The slope of this line is the response factor for the compound (Figure 1).

Figure 1. Example concentration curve. Sample (hypothetical) data values are shown in the table. The slope of the trend line is the response factor. Note: The y-intercept was used in the calculation of the trend line (0 mg/mL, 0 area).



Concentration (mg/mL) ⁴	Response (Area)
0.05	10,000
0.10	19,000
0.20	41,000

$$RF = 201,905$$

Once the response factor for a compound of interest is determined, calculation of the wt% of a sample can be done from the analysis of sample of known concentration using the following equation:

$$wt\% = \frac{A}{RF \times C_{cpd}} \times 100$$

Where,

- wt% = purity of the sample
- A = area of the peak of interest
- RF = response factor for the peak of interest (previously calculated)
- C_{cpd} = sample concentration of the compound being analyzed

References and Notes

¹ Pinciroli, V.; Biancardi, R.; Visentin, G.; Rizzo, V. "The Well-Characterized Synthetic Molecule: a Role for Quantitative ¹H NMR", *Org. Process Res. Dev.* **2004**, *8*, 381.

² It is more critical that >10 mg of compound and standard be used for the analysis than that the materials be used in an equimolar ratio. Errors in mass determination are the largest controllable source of error for this technique.

³ An acceptable chromatographic method should provide baseline separation of the peak of interest. The peak of interest should have a response within the linear range of the detector, have no shoulders or tailing, and should show good symmetry. This ensures that there is no variability in the response due to overlapping analyte peaks.

⁴ Standard concentrations should be corrected for the purity of the standard.

$$C = \frac{m \times P}{V}$$

Where,

- C is the concentration of the
- m is the measured mass of the standard sample
- P is the purity factor of the standard expressed as a decimal
- V is the volume of the diluted sample